

furnished a solid residue from which alkaloid 3 (8 mg), mp 224–225°, was obtained by prep. TLC on silica gel (butanone–xylene–Et₂NH–MeOH, 20:10:5:1) and repeated crystallization from EtOH. TLC (silica gel; solvent as above *R_f* 0.61) (Found C, 65.95, H, 6.50; N, 12.88. C₁₂H₁₄N₂O₂ requires: C, 66.02; H, 6.47; N, 12.84%).

Synthesis of (±)-1,2,3,9-tetrahydro-5-methoxy-pyrrolo[2,1-b]quinazoline-3-ol. 4-Amino-2-hydroxy-butyraldehyde diethyl-acetal (1.77 g) [7] was dissolved in H₂O (15 ml) and the pH of the soln adjusted to 2. The soln was then left at 80° for 30 min to liberate the free aldehyde (6). The pH of the soln of the aldehyde was adjusted to 5.5 with Pi buffer and the soln was added at 25° to a soln of 1.51 g 3-methoxyanthranilaldehyde (5) [8] in 50% (v/v) aq. MeOH (50 ml). The pH of the mixture was adjusted to 5.8 with Pi buffer and the mixture left at 25° for 3 days to give an orange soln of compound 7. The orange soln was then stirred vigorously at 60° in an atm of H₂ in the presence of 5% Pd–BaSO₄ catalyst for 1 hr. The mixture was filtered and the filtrate basified with NaOH and extracted with CHCl₃. The CHCl₃ extract was dried (Na₂SO₄) and the solvent evapd, when a light brown solid (1.2 g), mp 212–215°, was obtained. The solid

after three crystallizations from EtOH gave racemic 3 as colourless crystals (0.9 g), mp 224–225°. (Found C, 65.93; H, 6.52; N, 12.90. C₁₂H₁₄N₂O₂ requires: C, 66.02; H, 6.47; N, 12.84%.) Synthetic 3 was identical with the natural product (mp, mmp, TLC, IR, NMR).

REFERENCES

1. John, S., Groger, D. and Heese, M. (1971) *Helv. Chim. Acta.* **54**, 826 and references cited therein.
2. Amin, A. H., Mehta, D. R. and Samarth, S. S. (1970) *Prog. Drug Res.* **14**, 218.
3. Gupta, O. P., Anand, K. K., Ghattak Ray, B. J. and Atal, C. K. (1978) *Indian J. Exp. Biol.* **16**, 1075.
4. Arya, V. P. (1981) *Drugs of the Future* **6**, 373.
5. Bhalla, H. L. and Nimbkar, A. Y. (1982) *Drug Dev. Ind. Pharm.* **8**, 833.
6. Kuffner, F., Lenneis, G. and Bauer, H. (1960) *Monatsch. Chem.* **91**, 1152.
7. Leonard, N. J. and Martell, M. J. Jr. (1980) *Tetrahedron Letters* **44**.
8. Troger, J. and Dunker, E. (1925) *J. Prakt. Chem.* **111**, 207.

CYCLOBUXOVIRICINE, A STEROIDAL ALKALOID FROM *BUXUS PAPILOSA*

ATTA-UR-RAHMAN, M. IQBAL CHOUDHARY and (in part) MEHRUN NISA

H.E.J. Research Institute of Chemistry, University of Karachi, Karachi-32, Pakistan

(Revised received 22 March 1985)

Key Word Index—*Buxus papilosa*; Buxaceae; leaves; steroidal alkaloid; cyclobuxoviricine; ¹³C NMR.

Abstract—A new steroidal alkaloid has been isolated from the leaves of *Buxus papilosa*. Its structure has been assigned as 1 on the basis of spectroscopic studies.

INTRODUCTION

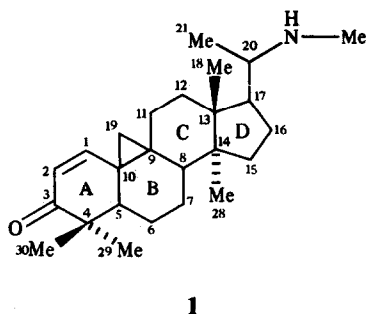
Buxus papilosa C. K. Schn, L. (Buxaceae) occurs abundantly in the northern regions of Pakistan. We have previously reported a number of new alkaloids from this plant [1, 2]. We now report the isolation and structural elucidation of a new steroidal alkaloid cyclobuxoviricine (1).

RESULTS AND DISCUSSION

Cyclobuxoviricine was isolated from the leaves of *B. papilosa* by extraction with ethanol, fractionation on the basis of differential basicity, CC and prep. TLC. Its IR spectrum showed bands at 1595 (C=C), 1647 (C=C–C=O)

[3] and 3350 cm^{−1} (NH). The UV spectrum was identical to that encountered in cyclobuxoviridine [4], showing maxima at 203 and 268 nm and a minimum at 230 nm.

The ¹H NMR spectrum (CDCl₃, 300 MHz) showed four singlets at δ 0.95, 0.90, 0.97 and 1.09, corresponding to the four tertiary methyl groups C-18, 28, 29, and 30, respectively. The secondary C-21 methyl group resonated as a doublet at δ 1.18 (*J*_{21,20} = 6.0 Hz), while the neighbouring C-20 methine proton appeared as a multiplet at δ 2.78 (*J*_{20,21} = 6 Hz, *J*_{20,17} = 9.8 Hz). Irradiation at δ 2.78 resulted in the collapse of the doublet of C-21 protons at δ 1.18 into a sharp singlet. A three-proton singlet resonated at δ 2.48, which was assigned to the –NMe group. A set of AB double doublets resonating at



1

Table 1. ^{13}C NMR spectral data of cyclobuxoviricine*

Carbon	Chemical shift (ppm)
1	153.18
2	127.08
3	199.20
4	45.53
5	49.20†
6	24.42
7	27.59
8	44.61
9	19.14‡
10	40.17
11	26.98
12	34.57
13	43.45
14	49.34†
15	31.94
16	29.02
17	49.99
18	16.50
19	19.50‡
20	58.57
21	18.60
28	17.15
29	23.40
30	21.50
NMe	29.90

*Recorded in CDCl_3 at 75 MHz.

†,‡ These values are interchangeable.

$\delta 0.75$ and 0.81 ($J_{19a, 19\beta} = 5.0$ Hz) was assigned to the cyclopropyl methylene protons. The downfield shift from the usual values [5] is due to the deshielding effect of the neighbouring olefinic functionality and has been observed earlier [3]. Two doublets at $\delta 6.76$ and 5.94 were ascribed to the C-1 and C-2 olefinic protons, respectively ($J_{1, 2} = 10.1$ Hz).

The ^{13}C NMR spectrum in CDCl_3 showed four signals at $\delta 16.5$, 17.15 , 21.5 , and 23.4 which were assigned to C-18, C-28, C-30 and C-29 methyl carbons, respectively. The C-2 olefinic carbon appeared at $\delta 127.08$, while the C-1 olefinic carbon β to the ketonic function resonated at $\delta 153.18$. The assignments to the various carbon atoms are shown in Table 1.

The mass spectrum of the compound afforded an $[\text{M}]^+$ at m/z 369.3025 which corresponds to the formula $\text{C}_{25}\text{H}_{39}\text{NO}$ (calc. 369.3031) indicating seven double bond equivalents. A peak at m/z 354.2759 ($\text{C}_{24}\text{H}_{36}\text{NO}$, calc. 354.2796) resulted from the loss of a methyl group from the $[\text{M}]^+$. Another peak at m/z 339.2551 was present corresponding to the loss of $-\text{NHMe}$ from the $[\text{M}]^+$, and this ion is commonly found in *Buxus* alkaloids bearing an $\text{Me}-\text{CHNHMe}$ side chain at C-17 [6]. A peak at m/z 312.2407 ($\text{C}_{22}\text{H}_{32}\text{O}$, calc. 312.2453) was consistent with the loss of $\text{C}_3\text{H}_7\text{N}^+$ from the $[\text{M}]^+$. The ion at m/z 85.0873 was in accordance with the composition $\text{C}_5\text{H}_{11}\text{N}$ (calc. 85.0891) which was attributed to cleavage of ring D [2]. The substance showed a base peak at m/z 57.0577 corresponding to $\text{C}_3\text{H}_7\text{N}^+$, which corresponds to the loss of $\text{Me}-\text{C}=\text{NH}-\text{Me}$ [1]. On the basis of these studies structure 1 is assigned to cyclobuxoviricine.

EXPERIMENTAL

MS were recorded on a double focussing spectrometer connected to a computer system. ^1H NMR spectra were recorded in CDCl_3 at 300 MHz, while ^{13}C NMR spectra were recorded at 75 MHz. The purity of the sample was checked on TLC (silica gel, Si F, precoated plates).

An EtOH extract of *B. papilosa* leaves (50 kg) collected from the northern regions of Pakistan, was evaporated to a gum. Total alkaloids were obtained by extraction into 10% HOAc. Partial separation of alkaloids was carried out by extraction into CHCl_3 at different pH values. The fraction obtained at pH 3.5 was loaded on a silica column (0.2–0.5 mm, 35–70 mesh ASTM). Elution with CHCl_3 –MeOH (9:1) afforded a number of close-moving alkaloids. The mixture was subjected to prep. TLC (silica gel) employing hexane– Me_2CO – Et_2NH (8:1.5:0.5), to afford

cyclobuxoviricine (17.5 mg) as a white amorphous solid, $[\alpha]_D^{10} = -54^\circ$ (CHCl_3). UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 203, 268, λ_{min} 230; IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 1595 ($\text{C}=\text{C}$), 1647 ($\text{C}=\text{C}-\text{C}=\text{O}$), 3350 (NH); ^1H NMR (300 MHz, CDCl_3): $\delta 0.75$ and 0.81 (2H, dd, $J_{19a, 19\beta} = 5.1$ Hz, C-19 CH_2), 0.90 (3H, s, 19-Me), 0.95 (3H, s, 18-Me), 0.97 (3H, s, 29-Me), 1.09 (3H, s, 30-Me), 2.48 (3H, s, NMe), 2.78 (1H, m, $J_{20, 21} = 6.0$ Hz, $J_{20, 17} = 9.8$ Hz, H-20), 5.94 (1H, d, $J_{2, 1} = 10.1$ Hz, H-2), 6.76 (1H, d, $J_{2, 1} = 10.1$ Hz, H-1); MS m/z (rel. int.): 369.3025 (72), 354.2759 (22), 339.2551 (14), 312.2407 (12), 85.0873 (61), 57.0577 (100).

REFERENCES

- Atta-ur-Rahman, Nisa, M. and Farhi, S. (1984) *Z. Naturforsch.* **39b**, 524.
- Atta-ur-Rahman, Nisa, M. and Jahan, K. (1985) *Phytochemistry* **24**, 1398.
- Nakano, T., Terao, S. and Saeki, Y. (1966) *J. Chem. Soc.* 1412.
- Julia, S., Julia, M., Tehen, S. Y. and Graffin, P. (1964) *Bull. Soc. Chim. France* 3207.
- Nakano, T. and Terao, S. (1965) *J. Chem. Soc.* 4512.
- Waller, G. R. and Deimes, O. C. (1980) *Biochemical Applications of Mass Spectroscopy*, p. 783. John Wiley, New York.